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# Liposidomycin C Inhibits Phospho-*N*-acetylmuramyl-pentapeptide Transferase in Peptidoglycan Synthesis of *Escherichia coli* Y-10

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Liposidomycin C ( $C_{42}H_{67}N_5O_{21}S$ , M.W. 1009) is a novel nucleoside antibiotic containing uracil, a sulfated aminosugar, and a fatty acid. It is a specific inhibitor of peptidoglycan synthesis of bacteria, inhibiting the formation of the lipid intermediates from uridine 5'-diphospho-*N*-acetylmuramyl-L-alanyl-D-glutamyl-meso-diaminopimelyl-D-[ $^{14}C$ ]alanyl-D-[ $^{14}C$ ]alanine and uridine 5'-diphospho-*N*-acetylglucosamine with a particulate enzyme from *Escherichia coli* Y-10. It also inhibited the formation of MurNAc(-pentapeptide)-P-P-lipid in the absence of UDP-GlcNAc. On the other hand, it inhibited the activity of *N*-acetylglucosamine transglycosylase and peptidoglycan transglycosylase only weakly using the same system from *E. coli*. Thus, it is concluded that the site of action of liposidomycin C is phospho-*N*-acetylmuramyl-pentapeptide transferase in peptidoglycan synthesis.

In screening for inhibitors of bacterial peptidoglycan synthesis, liposidomycins were found in the culture filtrate and mycelia of *Streptomyces griseosporeus*.<sup>1)</sup> These liposidomycins were found to contain at least twelve active components. Three major components, liposidomycins A, B, and C have been isolated and their physico-chemical properties report-

ed.<sup>1)</sup> Recently the structures of liposidomycins B and C were elucidated.<sup>2,3)</sup> They are novel lipid-containing uracil nucleosides of unusual complexity as shown in Fig. 1. Tunicamycin, a fatty acyl nucleoside antibiotic, also inhibited peptidoglycan synthesis and the site of action was reported to be the inhibition of the formation of lipid inter-

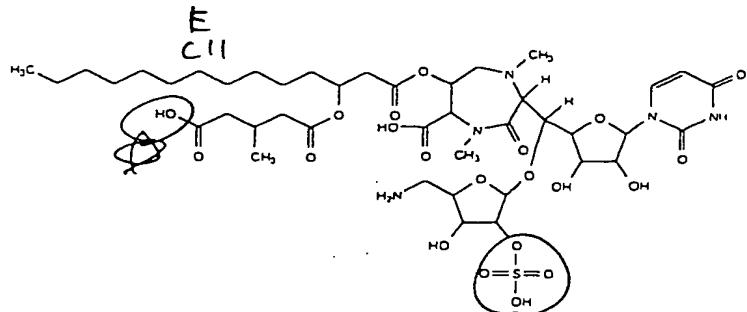


Fig. 1. Structure of Liposidomycin C.

*Abbreviations:* UDP-MurNAc-pentapeptide, Uridine 5'-diphospho-*N*-acetylmuramyl-L-alanyl-D-glutamyl-meso-diaminopimelyl-D-alanyl-D-alanine; UDP-GlcNAc, Uridine 5'-diphospho-*N*-acetylglucosamine.

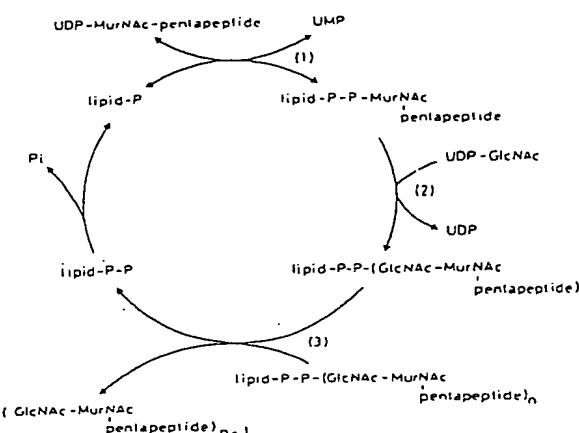


Fig. 2. Pathway of Peptidoglycan Synthesis from UDP-MurNAc-pentapeptide and UDP-GlcNAc with a Particulate Enzyme of *E. coli*.

(1) phospho-N-acetylmuramyl-pentapeptide transferase. (2) N-acetylglucosamine transglycosylase. (3) peptidoglycan transglycosylase. The pathway was the revised one from ref. 6.

mediates.<sup>4,5)</sup> It was found that liposidomycin C inhibited *in vitro* peptidoglycan synthesis of *E. coli* Y-10 having an activity with a magnitude three orders higher than that of tunicamycin.

This paper describes the site of action of liposidomycin C in peptidoglycan synthesis of *E. coli* Y-10. The primary site of action of liposidomycin C is found to be phospho-MurNAc-pentapeptide transferase, the first step of the lipid cycle of peptidoglycan synthesis in bacteria, as shown in Fig. 2.<sup>6)</sup>

### Materials and Methods

**Antibiotics.** Liposidomycin C was isolated as previously reported.<sup>11)</sup> Tunicamycin, vancomycin, and ristocetin were purchased from the Sigma Chemical Company, St. Louis, U.S.A. Enramycin was a gift from Takeda Chemical Industries, Ltd. Osaka, Japan.

**Radiochemicals.** UDP-[U-<sup>14</sup>C]GlcNAc (302 mCi/mmol) was purchased from Amersham. UDP-MurNAc-L-Ala-D-Glu-meso-Dap-D-[<sup>14</sup>C]Ala-D-[<sup>14</sup>C]Ala was prepared as previously described.<sup>7)</sup>

**Organisms and growth conditions.** *Bacillus cereus* T and *Escherichia coli* Y-10 were grown in bouillon medium (Eiken Chemical Co., Ltd.) at 37°C on a rotary shaker.

**Preparation of particulate enzyme of *E. coli* Y-10.** Particulate enzyme was prepared by grinding cells of *E. coli* Y-10 with sea sand (20–35 mesh) as previously reported.<sup>7)</sup>

**Preparation of UDP-MurNAc-L-Ala-D-Glu-meso-Dap-D-Ala-D-Ala.** UDP-MurNAc-pentapeptide was obtained after inducing its accumulation in cells of *B. cereus* T by treatment with 12.5 µg of vancomycin per ml as previously reported.<sup>7)</sup> From 10 l of the culture, approximately 600 OD units ( $A_{260}$ ) of UDP-MurNAc-pentapeptide were isolated.

**Assay of step 1 [phospho-MurNAc-pentapeptide transferase (EC 2.7.8.13)] in peptidoglycan synthesis from UDP-MurNAc-pentapeptide and UDP-GlcNAc with a particulate enzyme from *E. coli* Y-10.** The assay was done using a particulate enzyme prepared from *E. coli* Y-10 by a simple modification of the previous method.<sup>11)</sup> A reaction mixture (25 µl) containing 100 mM Tris-HCl (pH 7.5), 20 mM MgCl<sub>2</sub>, D-[<sup>14</sup>C]Ala-labeled UDP-MurNAc-pentapeptide (20,000 cpm), 0.1 mM UDP-GlcNAc (Sigma), 25 µl of antibiotic or distilled water, and 10 µl of particulate enzyme (protein concentration 15 mg/ml) was incubated for 10–60 min at 37°C. The reaction was stopped by the addition of 25 µl of 6 M pyridinium acetate (pH 4.2), and the lipid intermediates in the mixture were extracted twice with 100 µl of *n*-butanol.<sup>11)</sup> The extract was then transferred to a scintillation vial. The radioactivity was measured using a scintillation fluid, Pico-Fluor (Packard) with a liquid scintillation counter (lipid intermediates accumulation, step(s) 1 or 1 and 2).

The peptidoglycan that remained in the water layer was precipitated by adding an excess of 5% trichloroacetic acid. The precipitate was collected on a Whatman membrane filter GF/C, and washed twice with an excess of 5% trichloroacetic acid. The radioactivity of the precipitate on a filter was counted using a scintillation fluid, Filter-Count (Packard) with a liquid scintillation counter (peptidoglycan synthesis, steps 1, 2, and 3).

**Assay of step 2 [GlcNAc transglycosylase] in peptidoglycan synthesis from UDP-MurNAc-pentapeptide and UDP-GlcNAc with a particulate enzyme from *E. coli* Y-10.** A reaction mixture without UDP-GlcNAc (2.5 µl of 1 M Tris-HCl (pH 7.5), 2.5 µl of 0.2 M MgCl<sub>2</sub>, 5 µl of 2 mM UDP-MurNAc-pentapeptide, 5 µl of particulate enzyme, and 2.5 µl of distilled water) was incubated for 30 min at 37°C to accumulate MurNAc(-pentapeptide)-P-P-lipid, and then 5 µl of UDP-[U-<sup>14</sup>C]GlcNAc (20,000 cpm) and 2.5 µl of antibiotic or distilled water was added. After additional incubation of the reaction mixture for 10 min, lipid intermediates were assayed as described above.

**Assay of step 3 [peptidoglycan transglycosylase (EC 2.4.1.129)] in peptidoglycan synthesis from UDP-MurNAc-pentapeptide and UDP-GlcNAc with a particulate enzyme**

from *E. coli* Y-10. A reaction mixture (2.5  $\mu$ l of 1M Tris-HCl (pH 7.5), 2.5  $\mu$ l of 0.2M  $MgCl_2$ , 5  $\mu$ l of 2 mM UDP-MurNAc-pentapeptide, 5  $\mu$ l of UDP-[U- $^{14}C$ ]GlcNAc (20,000 cpm), 5  $\mu$ l of particulate enzyme, and 2.5  $\mu$ l of distilled water) was incubated for 5~20 min at 37°C to accumulate GlcNAc-MurNAc(-pentapeptide)-P-P-lipid. After incubation, 2.5  $\mu$ l of antibiotic or distilled water was added and the reaction mixture was incubated for 120 min at 37°C. The reaction mixture was extracted with *n*-butanol as described above and the water layer was spotted on Whatman 3MM paper. After ascending paper chromatography with isobutyric acid-1N  $NH_4OH$  (5:3), spots on the paper corresponding to peptidoglycan ( $R_f$  = 0) were cut out. The radioactivity was measured as described above.

## Results and Discussion

The effects of liposidomycin C on the formation of lipid intermediates and peptidoglycan have been examined with this system. The courses of lipid intermediate accumulation and peptidoglycan synthesis from D-[ $^{14}C$ ]Ala-labeled UDP-MurNAc pentapeptide and UDP-GlcNAc in the presence or the absence of liposidomycin C are shown in Fig. 3. With and without UDP-GlcNAc, formation of lipid intermediates and peptidoglycan were inhibited by liposidomycin C. This suggests

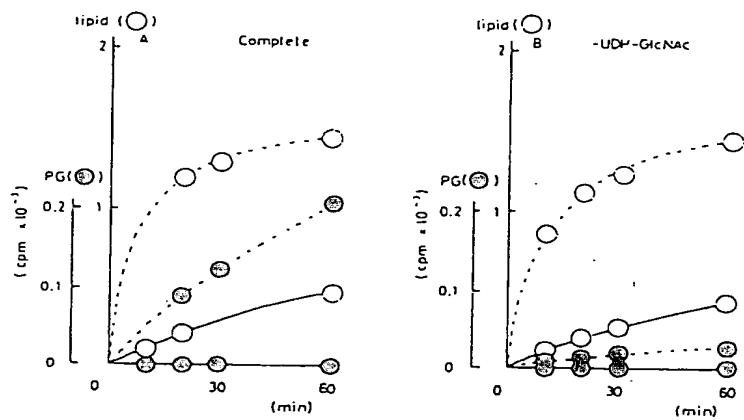


Fig. 3. Effects of Liposidomycin C on Lipid Intermediates and Peptidoglycan Syntheses by a Particulate Enzyme of *E. coli* Y-10.

Lipid intermediates and peptidoglycan were measured by the method described in Materials and Methods. Complete reaction mixture (A) and reaction mixture without UDP-GlcNAc (B).  $\circ$ , lipid intermediate;  $\bullet$ , peptidoglycan: -----, no antibiotic; —, liposidomycin C (1  $\mu$ g/ml).

Table I. EFFECTS OF LIPOSIDOMYCIN C ON PHOSPHO-N-ACETYL-MURAMYL-PENTAPEPTIDE TRANSFERASE ACTIVITY OF A PARTICULATE ENZYME OF *E. coli* Y-10

A reaction mixture (50  $\mu$ l) containing 100 mM Tris-HCl (pH 7.5), 20 mM  $MgCl_2$ , D-[ $^{14}C$ ]Ala-labeled UDP-MurNAc-pentapeptide (20,000 cpm), 0.1 mM UDP-GlcNAc, 5  $\mu$ l of liposidomycin C at various concentrations, and 5  $\mu$ l of particulate enzyme was incubated for 10 min at 37°C. Other methods are described in Materials and Methods.

Antibiotics	Concentration ( $\mu$ g/ml)	MurNAc(-pentapeptide)-P-P-lipid formation cpm (inhibition %)	
		- UDP-GlcNAc	+ UDP-GlcNAc
None	0	367 (0)	477 (0)
Liposidomycin C	0.025	241 (34)	333 (30)
	0.05	36 (90)	41 (91)
	0.1	17 (95)	31 (94)
	0.2	25 (93)	22 (95)
	1	21 (94)	9 (98)

Table II. EFFECTS OF LIPOSIDOMYCIN C ON *N*-ACETYLGLUCOSAMINE TRANSGLYCOSYLASE ACTIVITY OF A PARTICULATE ENZYME OF *E. coli* Y-10

A reaction mixture (2.5  $\mu$ l of 1 M Tris-HCl (pH 7.5), 2.5  $\mu$ l of 0.2 M  $MgCl_2$ , 5  $\mu$ l of 2 mM UDP-MurNAc-pentapeptide, 5  $\mu$ l of particulate enzyme, and 2.5  $\mu$ l of distilled water) was incubated for 30 min at 37°C (1st incubation). Then UDP-[U- $^{14}C$ ]GlcNAc (20,000 cpm) was added with and without antibiotic, and the reaction mixture was incubated for 10 min at 37°C (2nd incubation). Other methods are described in Materials and Methods.

Antibiotics	Concentration ( $\mu$ g/ml)	GluNAc-MurNAc(-pentapeptide)-P-P-lipid formation cpm (inhibition %)
None	0	1511 (0)
Liposidomycin C	0.1	1086 (28)
	1	921 (39)
Tunicamycin	10	946 (37)
	100	1101 (27)
Enramycin	500	419 (72)

that liposidomycin C inhibits step 1 thus inhibiting peptidoglycan synthesis.

Inhibitory effects of liposidomycin C at various concentration on phospho-MurNAc-pentapeptide transferase (step 1) were examined with and without UDP-GlcNAc. As shown in Table I, liposidomycin C inhibits 50% of phospho-MurNAc-pentapeptide transferase activity at 0.03  $\mu$ g/ml in both reactions. It also inhibits peptidoglycan synthesis at 0.038  $\mu$ g/ml which is comparable to phospho-MurNAc-pentapeptide transferase inhibition in the same system.

Next, we tested the effects of liposidomycin C on GlcNAc transglycosylase (step 2). After the formation of MurNAc(-pentapeptide)-P-P-lipid from UDP-MurNAc-pentapeptide in the absence of UDP-GlcNAc, liposidomycin C and the substrate (UDP-[ $^{14}C$ ]GlcNAc) were added. Formation of  $^{14}C$ -labeled GlcNAc-MurNAc(-pentapeptide)-P-P-lipid from MurNAc(-pentapeptide)-P-P-lipid and UDP-[ $^{14}C$ ]GlcNAc was completed after a 10-min incubation period. The butanol layer containing  $^{14}C$ -labeled GlcNAc-MurNAc(-pentapeptide)-P-P-lipid was then counted. Liposidomycin C did not inhibit step 2 strongly at 1  $\mu$ g/ml (39% inhibition), but enramycin (step 2 inhibitor)<sup>101</sup> showed 72% inhibition at 500  $\mu$ g/ml (Table II). Tunicamycin (step 1 inhibitor) also did not show inhibition in step 2 strongly at 100  $\mu$ g/ml (27% inhibition).

Table III. EFFECTS OF LIPOSIDOMYCIN C ON PEPTIDOGLYCAN TRANSGLYCOSYLASE ACTIVITY OF A PARTICULATE ENZYME OF *E. coli* Y-10

A reaction mixture (2.5  $\mu$ l of 1 M Tris-HCl (pH 7.5), 2.5  $\mu$ l of 0.2 M  $MgCl_2$ , 5  $\mu$ l of 2 mM UDP-MurNAc-pentapeptide, 5  $\mu$ l of UDP-[U- $^{14}C$ ]GlcNAc (20,000 cpm), 5  $\mu$ l of particulate enzyme, and 2.5  $\mu$ l of distilled water) was incubated for 5 min\* at 37°C (1st incubation). Then 2.5  $\mu$ l of antibiotic or distilled water was added, and the reaction mixture incubated for 2 hr at 37°C (2nd incubation). Other methods are described in Materials and Methods.

	Peptidoglycan synthesis cpm (inhibition %)
1st incubation	45
2nd incubation	
No antibiotic	364 (0)
Liposidomycin C (1 $\mu$ g/ml)	230 (42)
Vancomycin (100 $\mu$ g/ml)	70 (92)
Ristocetin (100 $\mu$ g/ml)	50 (98)

\* Similar results were obtained with 10 or 20 min of incubation.

We believe that the inhibition of over-all reactions is caused by the step 1 inhibition of liposidomycin C.

In the presence of enramycin (100  $\mu$ g/ml), lipid compounds (compound X) accumulated in the aqueous phase when the reaction mixture were extracted with *n*-butanol at pH 4.2.<sup>101</sup> But liposidomycin C (1  $\mu$ g/ml) and tu-

nicamycin (100  $\mu\text{g}/\text{ml}$ ) inhibited both water-soluble and butanol-soluble lipid intermediates (data not shown).

Next, we tested the effects of liposidomycin C on peptidoglycan transglycosylase. After the formation of GlcNAc-MurNAc(-pentapeptide)-P-P-lipid from UDP-MurNAc-pentapeptide and UDP-[ $^{14}\text{C}$ ]GlcNAc incubation (5 ~ 20 min), liposidomycin C was added and the formation of peptidoglycan was measured. Liposidomycin C slightly inhibited the peptidoglycan transglycosylase at 1  $\mu\text{g}/\text{ml}$  (30 ~ 40% inhibition). In contrast, the peptidoglycan transglycosylase inhibitors vancomycin<sup>11</sup> and ristocetin<sup>12</sup> were found to inhibit this reaction completely at 100  $\mu\text{g}/\text{ml}$  (90 ~ 100% inhibition, Table III). Vancomycin and ristocetin did not inhibit step 1 at 100  $\mu\text{g}/\text{ml}$  (data not shown).

It was already known that peptidoglycan transglycosylase was identical with the penicillin binding protein 1B.<sup>13,14</sup> Vancomycin and ristocetin were also inhibitors of penicillin binding protein 1B transglycosylase.

From the data described above, it is concluded that the primary site of action of liposidomycin C is phospho-*N*-acetylmuramyl-pentapeptide transferase (step 1), the first step of lipid cycle in peptidoglycan synthesis of bacteria. It is known that tunicamycin<sup>4,5</sup> and amphotomycin<sup>15</sup> also inhibit the same step, but liposidomycin C has been shown to be the most potent inhibitor (ID<sub>50</sub> of tunicamycin was about 12  $\mu\text{g}/\text{ml}$  in contrast to 0.038  $\mu\text{g}/\text{ml}$  for liposidomycin C in our conditions).<sup>4,15</sup> In contrast to the high *in vitro* activity, the anti-

bacterial activity of liposidomycin C is limited, as reported previously.<sup>11</sup> The reason has yet to be determined.

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